

Mitochondrial CO1 sequences of Banggai cardinal fish (BCF) from Lembeh Strait, North Sulawesi, Indonesia

Ari B. Rondonuwu, Lawrence J. L. Lumingas, Nego E. Bataragoa, Silvester B. Pratasik, Frans F. Tilaar, Meiske S. Salaki

Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115, North Sulawesi, Indonesia. Corresponding author: A. B. Rondonuwu, arirondonuwu@unsrat.ac.id

Abstract. This study determines the genetic structure of Banggai cardinal fish (BCF) from Lembeh Strait, North Sulawesi, based on the mitochondrial CO1 genetic marker. Sample collection used SCUBA gear and "chang net" fishing gear. One BCF specimen was collected in this study. Polymerase chain reaction (PCR) was performed using 5X Firepol PCR Master Mix. Bi-directional sequencing was done by First Base CO (Malaysia) using Big Dye© terminator chemistry (Perkin Elmer). PCR outcomes were separated using 0.8% agarose gel electrophoresis and shown by amplification of 718 bp band. The fish population found in the Lembeh Strait was *Pterapogon kauderni* Koumans, 1933 based on BLAST method. The lowest pairwise distance (d) was 0.000 (0%) and the highest was 0.324 (32.413%) with an average distance of 0.087 ± 0.117 (8.692±11.746%). Genetic distance (d) of the ingroup was shorter than that of the outgroup. Based on the phylogenetic tree, the population to *P. cauderni* is *Sphaeramia nematoptera*. **Key Words**: SCUBA, chang net, *P. kauderni, S. nematoptera*, genetic marker.

Introduction. One of the recently endangered living resources in Indonesian waters is coral reef fish of Apogonidae, *Pterapogon kauderni* Koumans, 1933 well-known as Banggai cardinal fish (BCF) and locally called *capungan* Banggai. As one of the seawater ornamental fish of high commercial value, the demand for this species is very high, despite the fact it is overfished in nature, particularly in its original habitat, Banggai islands covering Banggai regency and Banggai Laut regency. Recently, the species is in the status of endangered species of Red List of IUCN (International Union for the Conservation of Nature) due to exploitation and habitat degradation (Allen & Donaldson 2007). Since 2018, under the decree of Indonesian Marine Affairs and Fisheries Minister Number 49/KEPMEN-KP/2018, the status of limited protection of BCF has been established.

The occurrence of BCF outside the natural habitat is interesting to be studied because it was recently classified as an endemic fish. A species is called endemic if it is a native species found in certain place and not found in other area (Hugget 2004; Levin 2009). The area can be an island, country, or certain zone. Norman Myers of Oxford University used the number of endemic species as a criterion for identifying hotspots because it tends to have highly specific habitat or dietary requirements, low dispersal ability, and restricted geographical distributions (Russell et al 2017). Meanwhile, the distribution of this species has been detected in extensive regions outside Banggai group of island, such as Luwuk, Bali, and Lembeh Strait, North Sulawesi Province, in sufficiently high population (Erdmann & Vagelli 2001; Moore & Ndobe 2017; Vagelli 2008, 2011; Vagelli et al 2009).

AACL Bioflux, 2020, Volume 13, Issue 2. http://www.bioflux.com.ro/aacl

760

Commented [C1]: I add this word

Most biological experts think that major factor affecting speciation is geographic isolation, since as far as the population of the same species is still directly connecting or not, the gene flow still may occur (Sobel et al 2009). Nevertheless, species distribution can be controlled by geographic factors so that the exchange of gene structure in the population system and evolution will occur separately, and the evolution can make two populations be more different with time (Russell et al 2017).

Moreover, species identification of living creatures has developed, from morphological approach to molecular method. The latter used a short DNA part called "barcode DNA" that has applicative functions, such as ecological survey (Dick & Kress 2009), identification of cryptic taxa (Lahaye et al 2008), and confirmation of samples (Xue & Li 2011). As general understanding of barcode that can distinguish products, the DNA standard can ease the researcher to distinguish accurately and quickly the living species. Therefore, this study observes the biomolecular aspects of BCF in Lembeh Strait with the emphasis on the genetic characteristic and diversity (DNA barcode). This study was aimed at discerning the BCF from Lembeh Strait waters, North Sulawesi, using CO1 gene marker.

Material and Method

Sample collection. BCF samples were collected on July 5th, 2019 in Lembeh Strait waters, North Sulawesi, using SCUBA gear and "chang net" fishing gear, a modified beach seine facilitated with purse on the central part (Figure 1). The samples were preserved in 95% ethanol and stored at room temperature before DNA extraction.



Figure 1. Study site.

Sampling and preparation. About 3 g of BCF flesh was taken from the abdominal part. The flesh was preserved in 95% technical ethanol for at least 1 day before DNA extraction. The purpose of this preservation was to wash the sample from salt water and draw water from the cell so as to facilitate the DNA extraction process.

Extraction, PCR, and sequencing. Total DNA extraction (nucleus and mitochondria) of BCF employed innuPrep DNA Micro Kit (Analytical Jena, Germany). Polymerase chain

reaction (PCR) was performed using 5X Firepol PCR Master Mix. The CO1 gene (primary types) used are FF2d: 5'-TTC TCC ACC AAC CAC AAR GAY ATY GG-3' and FR1d 5'CAC CTC AGG GTG TCC GAA RAA YCA RAA-3' (Ivanova et al 2007). PCR was carried out in 35 cycles at 95°C (50 sec). The PCR product was visualized in 1% (b/v) agarose gel electrophoresis. Bi-directional sequencing was done by First Base CO (Malaysia) using Big Dye© terminator chemistry (Perkin Elmer).

Data analysis. The chromatogram obtained was edited using Mega X v10.1 software (Kumar et al 2018). Sequencing outcomes were compared with 8 other sequences of the same species, 4 sequences of different species of the same family Apogonidae, Cheilodipterus quinquelineatus, Apogon doederleini, Pristicon rhodopterus, and Sphaeramia nematoptera, and one butterflyfish Chaetodon kleinii (Chaetodontidae) from the Gen Bank using BLAST (Basic Local Alignment Search Tools) and BOLD systems (National Center for Biotechnology Information, U. s. National Library of Medicine. https://www.ncbi.nlm.nih.gov/genbank/). The sequence analysis of the BCF DNA was done by using p-distance model maximum composite likelihood (Tamura et al 2004), and phylogenetic analysis using Neighbour-Joining method (Saitou & Nei 1987).

Results and Discussion. The PCR outcomes were separated using 0.8% agarose gel electrophoresis and shown by amplification of 718 bp band (Figure 2). The DNA sequence of BCF from Lembeh Strait had genetic similarity to that of BCF (GenBank), especially sequences AB890097.1 and AP005997.1 with a total score of 1230 and query cover of 97%.



Figure 2. CO1 gene amplification of BCF samples from Lembeh Strait on 0.8% agarose gel (1kb: GeneRuler, Thermo Scientific as marker; AR1: sample number).

Based on species identity, all sequences of BCF had high similarity, 99.33-99.57%. The lowest similarity to BCF was found in that of the outgroup JF434784.1, the comparison with *Chaetodon kleinii*, with total cover of 606, 88% query, and 80.19% identity (Table 1). Thus, the fish sample collected from Lembeh Strait was *Pterapogon kauderni* Koumans, 1933 (Steinke et al 2009; Hubert et al 2012; Mabuchi et al 2014; Matias & Hereward 2018).

Table 1

Identification of banggai cardinal fish in Lembeh Strait using BLAST method (comparison with several sequences)

Sequence ID	Sequence	Total score	Query cover	<i>Identity</i> (%)	
AB890097.1	Pterapogon kauderni mitochondrial COI gene	1230	97	98.43	
	for cytochrome c oxidase subunit I				
AP005997.1	Pterapogon kauderni mitochondrial DNA,	1230	97	98.43	
	complete genome				
FJ583995.1	Pterapogon kauderni voucher BIOUG:HLC-10728	1179	90	99.54	
	cytochrome oxidase subunit 1 (COI) gene				
FJ583998.1	Pterapogon kauderni voucher BIOUG:HLC-10730	1173	90	99.38	
	cytochrome oxidase subunit 1 (COI) gene				
MH049167.1	Pterapogon kauderni isolate dki102 cytochrome	1099	86	98.33	
	c oxidase subunit 1 (COI) gene				
MH049167.1	Pterapogon kauderni isolate dki102 cytochrome	1079	83	99.33	
	c oxidase subunit 1 (COI) gene				
FJ346810.1	Pterapogon kauderni isolate A12 cytochrome	856	65	99.57	
	oxidase subunit I (COI) gene				
FJ346809.1	Pterapogon kauderni isolate A11 cytochrome	684	96	84.63	
	oxidase subunit I (COI) gene				
NC_040863.1	Cheilodipterus quinquelineatus mitochondrion	682	98	84.18	
AB890062.1	Apogon doederleini mitochondrial COI gene for	676	98	84.02	
	cytochrome c oxidase subunit I				
AB890095.1	Pristicon rhodopterus mitochondrial COI gene	667	98	83.85	
	for cytochrome c oxidase subunit I				
AB890106.1	Sphaeramia nematoptera mitochondrial COI	797	98	83.76	
	gene for cytochrome c oxidase subunit I				
JF434784.1	Chaetodon kleinii voucher REU0758 cytochrome	606	88	80.19	
	oxidase subunit 1 (COI) gene				

The sequence analysis of the 13 nucleotide sequences of 718 bp using the *maximum* composite likelihood model showed the lowest genetic distance (d) of 0.000 (0%) and the highest of 0.324 (32.413%) with mean population distance of 0.087 ± 0.117 (8.692±11.746%). The genetic distance (d) of the ingroup was shorter than that of the outgroup (Table 2 and Appendix 1).

Commented [C2]: When you put this as part of the text, I think it should be a Table?

Table 2 Average of pairwise distance of maximum composite likelihood model-based sequences

Sequences	P-distance (d)	%	SD
Total	0.087	8.692	0.117
Ingroup			
Pterapogon kauderni (7 populations)	0.001	0.140	0.002
Outgroup			
Apogonidae (4 populations)	0.199	19.901	0.001
Chaetodontidae (1 population)	0.324	32.413	

ix 1 Commented [C3]: Table 3?

No	Vo Sequence		2	3	4	5	6	7	8	9	10	11	12
1	Pterapogon kauderni_Lembeh_COI_AR1	0.000											
2	AB890097_Pterapogon kauderni_(Japan)	0.002	0.000										
3	AP005997_Pterapogon kauderni_(Japan)	0.000	0.002	0.000									
4	FJ583995_Pterapogon kauderni_(Canada)	0.000	0.002	0.000	0.000								
5	FJ583998_Pterapogon_kauderni_(Canada)	0.002	0.005	0.002	0.002	0.000							
6	MH049167_Pterapogon kauderni_(Indonesia)	0.005	0.007	0.005	0.005	0.007	0.000						
7	FJ346810_Pterapogon kauderni_(USA)	0.000	0.002	0.000	0.000	0.002	0.005	0.000					
8	FJ346809_Pterapogon kauderni_(USA)	0.000	0.002	0.000	0.000	0.002	0.005	0.000	0.000				
9	AB890106_Sphaeramia nematoptera_(Japan)	0.196	0.201	0.196	0.196	0.193	0.204	0.196	0.196	0.000			
10	NC_040863_Cheilodipterus quinquelineatus_(USA)	0.194	0.198	0.194	0.194	0.198	0.195	0.194	0.194	0.252	0.000		
11	AB890062_Apogon doederleini_(Japan)	0.192	0.188	0.192	0.192	0.196	0.192	0.192	0.192	0.169	0.180	0.000	
12	AB890095_Pristicon rhodopterus_(Japan)	0.213	0.218	0.213	0.213	0.217	0.214	0.213	0.213	0.194	0.195	0.180	0.000
13	JF434784 Chaetodon kleinii (France)	0.324	0.330	0.324	0.324	0.324	0.313	0.324	0.324	0.300	0.252	0.241	0.272

Appendix 1 Pair distances of banggai cardinal fish from Lembeh Strait with several populations - estimates of evolutionary divergence between sequences The pairwise distance method is the basis of the phylogenetic tree reconstruction. The value of p-distance index gives information on kinship among tree-like compared organisms. The more the p-distance value is, the farther the kinship between the organism, and vice versa. The genetic distance could reflect the genetic kinship between species, individuals or population. The present study indicates that the BCF population in Lembeh Strait has very close kinship to 7 other populations, although there is no information on the locality of population collected as ornamental fish.

Based on phylogenetic analysis using Neighbor-Joining method (Saitou & Nei 1987), the eight populations of BCF (ingroup) formed one cluster separated from other fish populations compared (outgroup) (Figure 3). According to Bernardi & Vagelli (2004), genetic isolation could occur in very short distance. However, the phylogenetic analysis of BCF in several natural habitats or the introduced habitat showed that BCF in Lembeh Strait belonged to the same clade as that in several localities of Banggai islands. Hoffman et al (2005) who studied the same species in Banggai islands and Luwuk waters using microsatelite method found no significant relationship between genetic and geographic distance despite increased genetic distance when removing Luwuk population from the analysis. Thus, to more accurately answer the genetic variations of BCF, more comprehensive phylogenetic studies need to be done with more populations from different locations.



Figure 3. The phylogenetic tree of Pterapogon kauderni.

The genetic distance analysis showed that four species of family Apogonidae, *Sphaeramia nematoptera*, *Pristicon rhodopterus*, *Cheilodipterus quinguelineatus* and *Apogon doederleini* are separated from *P. kauderni* population (Figure 3), but these four species occur in separate cluster. Besides Apogonidae cluster, Chaetodontidae population makes different cluster as well. This finding reveals that the kinship of fish species and family has

distinguishing genetic marker. *Chaetodon kleinii* is ecologically a fish that possesses the same habitat as *P. kauderni*, coral reef habitat (Allen & Erdmann 2012a, b; Kuiter & Tonozuka 2001; Sale 1991, 2002). BCF occupies most physiographic zones including reef flats, reef crests, and even the upper section of some reef slopes, but is limited to a narrow depth range of < 3 m, so that both species are grouped as coral fish (Vagelli 2011). In spite of that, the biological characteristics of both species are not mixed, such as inbreeding that will yield homozigotic individuals. It means that the biological character is a major factor determining the intraspecific genetic variations so that both species are always separate. Figure 3 also reveals that the population with the closest kinship to *P. kauderni* is *S. nematoptera*. In the revision of the systematics of the cardinal fishes (Percomorpha: Apogonidae) based on molecular analyses and comparative reevaluation of morphological characters, both species are classified as new tribe, Sphaeramiini (Mabuchi et al 2014).

Conclusions. The fish sample collected in Lembeh Strait was Banggai cardinal fish (BCF) *Pterapogon kauderni* Koumans, 1933. The genetic distance (d) of the ingroup is shorter than that of the outgroup. The BCF population in Lembeh Strait and 7 other BCF populations had very close kinship. In the phylogenetic tree, the eight populations formed one cluster that was separated from the other species compared and outgroup. Other population having the closest kinship to *P. kauderni* was *S. nematoptera*.

References

- Allen G. R, Donaldson T. J., 2007 *Pterapogon kauderni*. The IUCN red list of threatened species 2007:e.T63572A12692964.
- Allen G. R., Erdmann M. V., 2012a Reef fishes of the East Indies. Volume I. Tropical Reef Research, Perth, Australia, 424 pp.
- Allen G. R., Erdmann M. V. 2012b Reef fishes of the East Indies. Volume II. Tropical Reef Research, Perth, Australia, 856 pp.
- Bernardi G., Vagelli A., 2004 Population structure in Banggai cardinal fish, *Pterapogon kauderni*, a coral reef species lacking a pelagic larval phase. Marine Biology 145:803-810.
- Decree of the Ministry of Marine Affairs and Fisheries of the Republic of Indonesia No. 49/KepMen-KP/2018 on the determination of the limited protection status of banggai cardinal fish (*Pterapogon kauderni*), 8 pp.
- Dick C. W., Kress W. J., 2009 Dissecting tropical plant diversity with forest plots and a molecular toolkit. BioScience 59(9):745-755.
- Erdmann M., Vagelli A., 2001 Banggai cardinal fish invade Lembeh Strait. Coral Reefs 20:252-253.
- Hoffman E. A., Kolm N., Berglund A., Arguello J. R., Jonees A. G., 2005 Genetic structure in the coral-reef-associated Banggai cardinal fish, *Pterapogon kauderni*. Molecular Ecology 14:1367-1375.
- Hubert N., Meyer C. P., Bruggemann H. J., Guerin F., Komeno R. J., Espiau B., Causse R., Williams J. T., Planes S., 2012 Cryptic diversity in Indo-Pacific coral-reef fishes revealed by DNA-barcoding provides new support to the centre-of-overlap hypothesis. PLoS ONE 7(3):e28987.

Hugget R. J., 2004 Fundamentals of biogeography. 2nd edition, Routledge, UK, 456 pp.

- Ivanova N. V., Zemlak T. S., Hanner R. H., Hebert P. D. N., 2007 Universal primer cocktails for fish DNA barcoding. Molecular Ecology Notes 7(4):544-548.
- Kuiter R. H., Tonozuka T., 2001 Pictorial guide to Indonesian reef Fishes Part 2. Zoonetics, Australia, 622 pp.
- Kumar S., Stecher G., Li M., Knyaz C., Tamura K., 2018 MEGA X: molecular evolutionary genetics analysis across computing platforms. Molecular Biology and Evolution 35(6):1547-1549.
- Lahaye R., van der Bank M., Bogarin D., Warner J., Pupulin F., Gigot G., Maurin O., Duthoit S., Barraclough T. G., Savolainen V., 2008 DNA barcoding the floras of biodiversity hotspots. Proceedings of the National Academy of Sciences of the USA 105(8):2923-2928.

Levin S. A., 2009 The Princeton guide to ecology. Princeton University Press, New Jersey, 848 pp.

- Mabuchi K., Fraser T. H., Song H., Azuma Y., Nishida M., 2014 Revision of the systematics of the cardinal fishes (Percomorpha: Apogonidae) based on molecular analyses and comparative reevaluation of morphological characters. Zootaxa 3846(2):151-203.
- Matias A. M., Hereward J., 2018 The complete mitochondrial genome of the five-lined cardinal fish *Cheilodipterus quinquelineatus* (Apogonidae). Mitochodrial DNA Part B 3(2):521-522.
- Moore A. M., Ndobe S., Jompa J., 2017 [A site-based conservation approach to promote the recovery of Banggai cardinal fish (*Pterapogon kauderni*) endemic populations]. Coastal and Ocean Journal 1(2):63-72. [in Indonesian]
- Russell P. J., Hertz P. E., McMillan B., 2017 Biology the dynamic science. 4th edition, Cengage Learning, Boston, USA, 1456 pp.
- Saitou N., Nei M., 1987 The neighbor-joining method: a new method for reconstructing phylogenetic trees. Molecular Biology and Evolution 4(4):406-425.
- Sale P. F., 1991 The ecology of fishes on coral reefs. Academic Press, Inc., San Diego, California, 754 pp.
- Sale P. F., 2002 Coral reef fishes dynamics and diversity in a complex ecosystem. Elsevier Sciences, 549 pp.
- Sobel J. M., Chen G. F., Watt L. R., Schemske D. W., 2009 The biology of speciation. Evolution 64(2):295-315.
- Steinke D., Zemlak T. S., Hebert P. D. N., 2009 Barcoding Nemo: DNA-based identifications for the ornamental fish trade. PLoS ONE 4(7):e6300.
- Tamura K., Nei M., Kumar S., 2004 Prospects for inferring very large phylogenies by using the neighbor-joining method. Proceedings of the National Academy of Sciences of the USA 101(30):11030-11035.
- Vagelli A. A., 2008 The unfortunate journey of *Pterapogon kauderni*: a remarkable apogonid endangered by the international ornamental fish trade, and its case in CITES. SPC Live Reef Fish Information Bulletin 18:17-28.
- Vagelli A. A., 2011 The Banggai cardinal fish: natural history, conservation, and culture of *Pterapogon kauderni*. Wiley-Blackwell, UK, 224 pp.
- Vagelli A., Burford M., Bernardi G., 2009 Fine scale dispersal in Banggai cardinal fish, *Pterapogon kauderni,* a coral reef species lacking a pelagic larval phase. Marine Genomics 1(3-4):129-134.
- Xue C. Y., Li D. Z., 2011 Use of DNA barcode sensu lato to identify traditional Tibetan medicinal plant *Gentianopsis paludosa* (Gentianaceae). Journal of Systematics and Evollution 49(3):267-270.

Received: 01 March 2020. Accepted: 27 March 2020. Published online: 03 April 2020. Authors:

Ari B. Rondonuwu, Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115, North Sulawesi, Indonesia, e-mail: arirondonuwu@unsrat.ac.id.

- Lawrence J. L. Lumingas, Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu,
- Manado-95115, North Sulawesi, Indonesia, e-mail: ljllumingas@yahoo.com

Nego E. Bataragoa, Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115, North Sulawesi, Indonesia, e-mail: nebgoa@unsrat.ac.id

Silvester B. Pratasik, Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115, North Sulawesi, Indonesia, e-mail: spjong07@yahoo.com

Frans F. Tilaar, Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115, North Sulawesi, Indonesia, e-mail: fftilaar@unsrat.co.id

Meiske S. Salaki, Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115, North Sulawesi, Indonesia, e-mail: mssalaki@unsrat.ac.id

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

How to cite this article:

Rondonuwu A. B., Lumingas L. J. L., Bataragoa N. E., Pratasik S. B., Tilaar F. F., Salaki M. S., 2020 Mitochondrial CO1 sequences of banggai cardinal fish (BCF) from Lembeh Strait, North Sulawesi, Indonesia. AACL Bioflux 13(2):760-767.