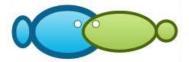


Gambar 3. Surat Hasil Review



Mitochondrial CO1 sequences of Banggai cardinal fish (BCF) from Lembeh Strait, North Sulawesi, Indonesia

Ari B. Rondonuwu, Lawrence J. L. Lumingas, Nego E. Bataragoa, Silvester B. Pratasik, Frans F. Tilaar, Meiske S. Salaki

Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115, North Sulawesi, Indonesia. Corresponding author: A. B. Rondonuwu, arirondonuwu@unsrat.ac.id

Abstract. This study determines Banggai cardinal fish (BCF) from Lembeh Strait, North Sulawesi, based on the mitochondrial CO1 genetic marker. Sample collection used SCUBA gear and "chang net" fishing gear. One BCF specimen was collected in this study. Polymerase chain reaction (PCR) was performed using 5X Firepol PCR Master Mix. Bi-directional sequencing was done by First Base CO (Malaysia) using Big Dye⊚ terminator chemistry (Perkin Elmer). PCR outcomes were separated using 0.8% agarose gel electrophoresis and shown by amplification of 718 bp band. The fish population found in the Lembeh Strait was *Pterapogon kauderni* Koumans, 1933 based on BLAST method. The lowest pairwise distance (d) was 0.000 (0%) and the highest was 0.324 (32.413%) with an average distance of 0.087±0.117 (8.692±11.746%). Genetic distance (d) of the ingroup was shorter than that of the outgroup. Based on the phylogenetic tree, the population of *P. cauderni* (ingroup) forms one cluster and is separated from other fish groups (outgroup). The closest population to *P. kauderni* is *Sphaeramia nematoptera*.

Key Words: SCUBA, chang net, P. kauderni, S. nematoptera, genetic marker.

Introduction. One of the recently endangered living resources in Indonesian waters is coral reef fish of Apogonidae, *Pterapogon kauderni* Koumans, 1933 well-known as Banggai cardinal fish (BCF) and locally called *capungan* Banggai. As one of the seawater ornamental fish of high commercial value, the demand for this species is very high, despite the fact it is overfished in nature, particularly in its original habitat, Banggai islands covering Banggai regency and Banggai Laut regency. Recently, the species is in the status of endangered species of Red List of IUCN (International Union for the Conservation of Nature) due to exploitation and habitat degradation (Allen & Donaldson 2007). Since 2018, under the decree of Indonesian Marine Affairs and Fisheries Minister Number 49/KEPMEN-KP/2018, the status of limited protection of BCF has been established.

The occurrence of BCF outside the natural habitat is interesting to be studied because it was recently classified as an endemic fish. A species is called endemic if it is a native species found in certain place and not found in other area (Hugget 2004; Levin 2009). The area can be an island, country, or certain zone. Norman Myers of Oxford University used the number of endemic species as a criterion for identifying hotspots because it tends to have highly specific habitat or dietary requirements, low dispersal ability, and restricted geographical distributions (Russell et al 2017). Meanwhile, the distribution of this species has been detected in extensive regions outside Banggai group of island, such as Luwuk, Bali, and Lembeh Strait, North Sulawesi Province, in sufficiently population (Erdmann & Vagelli 2001; Moore & Ndobe 2017; Vagelli 2008, 2011; Vagelli et al 2009).

Commented [indra1]: determines what? The identification of BFC?

Commented [indra2]: to be put on the references list as well

Commented [indra3]: be more specific here

Most biological experts think that major factor affecting speciation is geographic isolation, since as far as the population of the same species is still directly connecting or not, the gene flow still may occur (Sobel et al 2009). Nevertheless, species distribution can be controlled by geographic factors so that the exchange of gene structure in the population system and evolution will occur separately, and the evolution can make two populations be more different with time (Russell et al 2017).

Moreover, species identification of living creatures has developed, from morphological approach to molecular method. The latter used a short DNA part called "barcode DNA" that has applicative functions, such as ecological survey (Dick & Kress 2009), identification of cryptic taxa (Lahaye et al 2008), and confirmation of samples (Xue & Li 2011). As general understanding of barcode that can distinguish products, the DNA standard can ease the researcher to distinguish accurately and quickly the living species. Therefore, this study observes the biomolecular aspects of BCF in Lembeh Strait with the emphasis on the genetic characteristic and diversity (DNA barcode). This study was aimed at discerning the BCF from Lembeh Strait waters, North Sulawesi, using CO1 gene marker.

Material and Method

Sample collection. BCF samples were collected in Lembeh Strait waters, North Sulawesi, using SCUBA gear and "chang net" fishing gear, a modified beach seine facilitated with purse on the central part. The samples were preserved in 95% ethanol and stored at room temperature before DNA extraction.

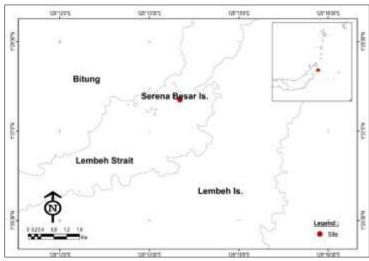


Figure 1. Study site.

Sampling and preparation. About 3 g of fish flesh was taken from the abdominal part. It was preserved in 95% technical ethanol for at least 1 day before extraction. The purpose of this preservation is to wash the sample from salt water and draw water from the cell so as to facilitate the DNA extraction process.

Extraction, PCR, and sequencing. Total DNA extraction (nucleus and mitochondria) of BCF employed innuPrep DNA Micro Kit (Analytical Jena, Germany). Polymerase chain

Commented [indra4]: when?

Commented [indra5]: quote the figure in text too

Commented [indra6]: who was preserved? The entire fish? Or the flesh only, prior to DNA extraction?

reaction (PCR) was performed using 5X Firepol PCR Master Mix. The CO1 gene (primary types) used are FF2d: 5'-TTC TCC ACC AAC CAC AAR GAY ATY GG-3' and FR1d 5'CAC CTC AGG GTG TCC GAA RAA YCA RAA-3' (Ivanova et al 2007). PCR was carried out in 35 cycles at 95°C (50 sec). The PCR product was visualized in 1% (b/v) agarose gel electrophoresis. Bi-directional sequencing was done by First Base CO (Malaysia) using Big Dye© terminator chemistry (Perkin Elmer).

Data analysis. The chromatogram obtained was edited using Mega X v10.1 software (Kumar et al 2018). Sequencing outcomes were compared with several sequences of the same species, different species of the same family (Apogonidae) and one butterflyfish species (Chaetodontidae) from the Gen Bank using BLAST (Basic Local Alignment Search Tools) and BOLD systems (National Center for Biotechnology Information, U.S. National Library of Medicine; **Error! Hyperlink reference not valid.**).

Results and Discussion. The PCR outcomes were separated using 0.8% agarose gel electrophoresis and shown by amplification of 718 bp band (Figure 2). The DNA sequence of BCF from Lembeh Strait had genetic similarity to that of BCF (GenBank), especially sequences AB890097.1 and AP005997.1 with a total score of 1230 and query cover of 97%.



Figure 2. CO1 gene amplification of BCF samples from Lembeh Strait on 0.8% agarose gel (1kb: GeneRuler, Thermo Scientific as marker; AR1: sample number).

Based on species identity, all sequences of BCF had high similarity, 99.33-99.57%. The lowest similarity to BCF was found in that of the outgroup JF434784.1, the comparison with *Chaetodon kleinii*, with total cover of 606, 88% query, and 80.19% identity. Thus, the fish sample collected from Lembeh Strait was *Pterapogon kauderni* Koumans, 1933 (Steinke et al 2009; Hubert et al 2012; Mabuchi et al 2014; Matias & Hereward 2018).

Commented [indra7]: how many sequences?

Commented [indra8]: add their names

Commented [indra9]: provide the scientific name

Commented [indra10]: ???

Commented [indra11]: we are sorry, but the figure is unreadable in this form. Try to provide a much better one. And, if possible, lay it down horizontally.

Table 1

Identification of banggai cardinal fish in Lembeh Strait using BLAST method (comparison with several sequences)

Commented [indra12]: we cannot see the table's quotation in text...

Sequence ID	Sequen c e	Total score	Query cover	Identity (%)
AB890097.1	Pterapogon kauderni mitochondrial COI gene	1230	97	98.43
AB030037.1	for cytochrome c oxidase subunit I	1230	5,	50.45
AP005997.1	Pterapogon kauderni mitochondrial DNA,	1230	97	98.43
	complete genome			
FJ583995.1	Pterapogon kauderni voucher BIOUG:HLC-10728	1179	90	99.54
F1F02000 4	cytochrome oxidase subunit 1 (COI) gene	4470	00	00.00
FJ583998.1	Pterapogon kauderni voucher BIOUG:HLC-10730	1173	90	99.38
MH049167.1	cytochrome oxidase subunit 1 (COI) gene Pterapogon kauderni isolate dki102 cytochrome	1099	86	98.33
МП049167.1	c oxidase subunit 1 (COI) gene	1099	00	90.33
MH049167.1	Pterapogon kauderni isolate dki102 cytochrome	1079	83	99.33
1110 15107.1	c oxidase subunit 1 (COI) gene	1075	- 03	JJ.33
FJ346810.1	Pterapogon kauderni isolate A12 cytochrome	856	65	99.57
	oxidase subunit I (COI) gene			
FJ346809.1	Pterapogon kauderni isolate A11 cytochrome	684	96	84.63
	oxidase subunit I (COI) gene			
NC_040863.1	Cheilodipterus quinquelineatus mitochondrion	682	98	84.18
AB890062.1	Apogon doederleini mitochondrial COI gene for	676	98	84.02
	cytochrome c oxidase subunit I			
AB890095.1	Pristicon rhodopterus mitochondrial COI gene	667	98	83.85
	for cytochrome c oxidase subunit I			
AB890106.1	Sphaeramia nematoptera mitochondrial COI	797	98	83.76
	gene for cytochrome c oxidase subunit I			00.40
JF434784.1	Chaetodon kleinii voucher REU0758 cytochrome	606	88	80.19
	oxidase subunit 1 (COI) gene			

Commented [indra13]: why are these two sequences underlined?

Commented [indra14]: why are these two sequences underlined?

The sequence analysis of the BCF DNA of 718 bp using p-distance model maximum composite likelihood (Tamura et al 2004) showed the lowest genetic distance (d) of 0.000 (0%) and the highest of 0.324 (32.413%) with mean population distance of 0.087 ± 0.117 (8.692 $\pm11.746\%$). The genetic distance (d) of the ingroup was shorter than that of the outgroup (Table 2 and Appendix 1).

Table 2

Average of pairwise distance several populations (model maximum composite likelihood)

	P-distance (d)	%	SD
Total	0.087	8.692	0.117
Ingroup			
Pterapogon kauderni (7 populations)	0.001	0.140	0.002
Outgroup			
Apogonidae (4 populations)	0.199	19.901	0.001
Chaetodontidae (1 population)	0.324	32.413	

Commented [indra15]: ??? make it more clear

Commented [indra16]: what do we have in this column?

Appendix 1 Pair distances of banggai cardinal fish from Lembeh Strait with several populations - estimates of evolutionary divergence between sequences

No	Sequence	1	2	3	4	5	6	7	8	9	10	11	12
1	Pterapogon kauderni_Lembeh_COI_AR1	0.000											
2	AB890097_Pterapogon kauderni_(Japan)	0.002	0.000										
3	AP005997_Pterapogon kauderni_(Japan)	0.000	0.002	0.000									
4	FJ583995_Pterapogon kauderni_(Canada)	0.000	0.002	0.000	0.000								
5	FJ583998_Pterapogon_kauderni_(Canada)	0.002	0.005	0.002	0.002	0.000							
6	MH049167_Pterapogon kauderni_(Indonesia)	0.005	0.007	0.005	0.005	0.007	0.000						
7	FJ346810_Pterapogon kauderni_(USA)	0.000	0.002	0.000	0.000	0.002	0.005	0.000					
8	FJ346809_Pterapogon kauderni_(USA)	0.000	0.002	0.000	0.000	0.002	0.005	0.000	0.000				
9	AB890106_Sphaeramia nematoptera_(Japan)	0.196	0.201	0.196	0.196	0.193	0.204	0.196	0.196	0.000			
10	NC_040863_Cheilodipterus quinquelineatus_(USA)	0.194	0.198	0.194	0.194	0.198	0.195	0.194	0.194	0.252	0.000		
11	AB890062_Apogon doederleini_(Japan)	0.192	0.188	0.192	0.192	0.196	0.192	0.192	0.192	0.169	0.180	0.000	
12	AB890095_Pristicon rhodopterus_(Japan)	0.213	0.218	0.213	0.213	0.217	0.214	0.213	0.213	0.194	0.195	0.180	0.000
13	JF434784_Chaetodon kleinii_(France)	0.324	0.330	0.324	0.324	0.324	0.313	0.324	0.324	0.300	0.252	0.241	0.272

The number of base substitutions per site from between sequences are shown. Analyses were conducted using the Maximum Composite Likelihood model [1]. This analysis involved 13 nucleotide sequences. Codon positions included were 1st+2nd+3rd+noncoding. All positions containing gaps and missing data were eliminated (complete deletion option). There were a total of 409 positions in the final dataset. Evolutionary analyses were conducted in MEGA X [2].

Commented [indra17]: check the grammar

Commented [indra18]: what does this mean?

Commented [indra19]: what does this mean?

The pairwise distance method is the basis of the phylogenetic tree reconstruction. The value of p-distance index gives information on kindship among tree-like compared organisms. The more the p-distance value is, the farther the kinship between the organism, and vice versa. The genetic distance could reflect the genetic kinship between species, individuals or population. The present study indicated that the BCF population in Lembeh Strait had very close kinship to 7 other populations, although there is no information on the locality of population collected as ornamental fish.

Based on phylogenetic analysis using Neighbor-Joining method (Saitou & Nei 1987), the eight populations of BCF (ingroup) formed one cluster separated from other fish populations compared (outgroup) (Figure 2). According to Bernardi & Vagelli (2004), genetic isolation could occur in very short distance. However, the phylogenetic analysis of BCF in several natural habitats or the introduced habitat showed that BCF in Lembeh Strait belonged to the same clade as that in several localities of Banggai islands. Hoffman et al (2005) who studied the same species in Banggai islands and Luwuk waters using microsatelite method found no significant relationship between genetic and geographic distance despite increased genetic distance when removing Luwuk population from the analysis. Thus, to more accurately answer the genetic variations of BCF, more comprehensive phylogenetic studies need to be done with more populations from different locations.

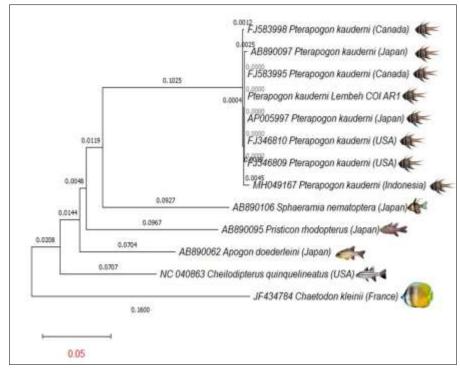


Figure 2. The phylogenetic tree of Pterapogon kauderni.

Commented [indra20]: actually it should be kinship (without d

The genetic distance analysis showed that four species of family Apogonidae, Sphaeramia nematoptera, Pristicon rhodopterus, Cheilodipterus quinguelineatus and Apogon doederleini are separated from P. kauderni population (Figure 2), but these four species occur in separate cluster. Besides Apogonidae cluster, Chaetodontidae population makes different cluster as well. This finding reveals that the kinship of fish species and family has distinguishing genetic marker. C. kleinii is ecologically fish group that possesses the same habitat as P. kauderni, coral reef habitat, so that both species are grouped as coral fish. In spite of that, the biological characteristics of both species are not mixed, such as inbreeding that will yield homozigotic individuals. It means that the biological character is a major factor determining the intraspecific genetic variations so that both species are always separate. Figure 2 also reveals that the population with the closest kinship to P. kauderni is S. nematoptera. In the revision of the systematics of the cardinal fishes (Percomorpha: Apogonidae) based on molecular analyses and comparative reevaluation of morphological characters, both species are classified as new tribe, Sphaeramiini (Mabuchi et al 2014).

Conclusions. The fish sample collected in Lembeh Strait was Banggai cardinal fish (BCF) *Pterapogon kauderni* Koumans, 1933. The genetic distance (d) of the ingroup is shorter than that of the outgroup. The BCF population in Lembeh Strait and 7 other BCF populations had very close kinship. In the phylogenetic tree, the eight populations formed one cluster that was separated from the other species compared and outgroup. Other population having the closest kinship to *P. kauderni* was *S. nematoptera*.

References

- Allen G. R, Donaldson T. J., 2007 *Pterapogon kauderni*. The IUCN red list of threatened species 2007:e.T63572A12692964.
- Bernardi G., Vagelli A., 2004 Population structure in Banggai cardinal fish, *Pterapogon kauderni*, a coral reef species lacking a pelagic larval phase. Marine Biology 145:803-810.
- Dick C. W., Kress W. J., 2009 Dissecting tropical plant diversity with forest plots and a molecular toolkit. BioScience 59(9):745-755.
- Erdmann M., Vagelli A., 2001 Banggai cardinal fish invade Lembeh Strait. Coral Reefs 20:252-253.
- Hoffman E. A., Kolm N., Berglund A., Arguello J. R., Jonees A. G., 2005 Genetic structure in the coral-reef-associated Banggai cardinal fish, *Pterapogon kauderni*. Molecular Ecology 14:1367-1375.
- Hubert N., Meyer C. P., Bruggemann H. J., Guerin F., Komeno R. J., Espiau B., Causse R., Williams J. T., Planes S., 2012 Cryptic diversity in Indo-Pacific coral-reef fishes revealed by DNA-barcoding provides new support to the centre-of-overlap hypothesis. PLoS ONE 7(3):e28987.
- Hugget R. J., 2004 Fundamentals of biogeography. 2nd edition, Routledge, UK, 456 pp.
- Ivanova N. V., Zemlak T. S., Hanner R. H., Hebert P. D. N., 2007 Universal primer cocktails for fish DNA barcoding. Molecular Ecology Notes 7(4):544-548.
- Kumar S., Stecher G., Li M., Knyaz C., Tamura K., 2018 MEGA X: molecular evolutionary genetics analysis across computing platforms. Molecular Biology and Evolution 35(6):1547-1549.
- Lahaye R., van der Bank M., Bogarin D., Warner J., Pupulin F., Gigot G., Maurin O., Duthoit S., Barraclough T. G., Savolainen V., 2008 DNA barcoding the floras of biodiversity hotspots. Proceedings of the National Academy of Sciences of the USA 105(8):2923-2928.
- Levin S. A., 2009 The Princeton guide to ecology. Princeton University Press, New Jersey, 848 pp.

Commented [indra21]: add references here

- Mabuchi K., Fraser T. H., Song H., Azuma Y., Nishida M., 2014 Revision of the systematics of the cardinal fishes (Percomorpha: Apogonidae) based on molecular analyses and comparative reevaluation of morphological characters. Zootaxa 3846(2):151-203.
- Matias A. M., Hereward J., 2018 The complete mitochondrial genome of the five-lined cardinal fish Cheilodipterus quinquelineatus (Apogonidae). Mitochodrial DNA Part B 3(2):521-522.
- Moore A. M., Ndobe S., Jompa J., 2017 [A site-based conservation approach to promote the recovery of Banggai cardinal fish (Pterapogon kauderni) endemic populations]. Coastal and Ocean Journal 1(2):63-72. [in Indonesian]
- Russell P. J., Hertz P. E., McMillan B., 2017 Biology the dynamic science. 4th edition, Cengage Learning, Boston, USA, 1456 pp.
- Saitou N., Nei M., 1987 The neighbor-joining method: a new method for reconstructing phylogenetic trees. Molecular Biology and Evolution 4(4):406-425.
- Sobel J. M., Chen G. F., Watt L. R., Schemske D. W., 2009 The biology of speciation. Evolution 64(2):295-315.
- Steinke D., Zemlak T. S., Hebert P. D. N., 2009 Barcoding Nemo: DNA-based identifications for the ornamental fish trade. PLoS ONE 4(7):e6300.
- Tamura K., Nei M., Kumar S., 2004 Prospects for inferring very large phylogenies by using the neighbor-joining method. Proceedings of the National Academy of Sciences of the USA 101(30):11030-11035.
- Vagelli A. A., 2008 The unfortunate journey of Pterapogon kauderni: a remarkable apogonid endangered by the international ornamental fish trade, and its case in CITES. SPC Live Reef Fish Information Bulletin 18:17-28.
- Vagelli A. A., 2011 The Banggai cardinal fish: natural history, conservation, and culture of Pterapogon kauderni. Wiley-Blackwell, UK, 224 pp.
- Vagelli A., Burford M., Bernardi G., 2009 Fine scale dispersal in Banggai cardinal fish, Pterapogon kauderni, a coral reef species lacking a pelagic larval phase. Marine Genomics 1(3-4):129-134.
- Xue C. Y., Li D. Z., 2011 Use of DNA barcode sensu lato to identify traditional Tibetan medicinal plant Gentianopsis paludosa (Gentianaceae). Journal of Systematics and Evollution 49(3):267-270.

Received: 01 March 2020, Accepted: 27 March 2020, Published online: xx April 2020,

Authors:

Ari B. Rondonuwu, Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115, North Sulawesi, Indonesia, e-mail: arirondonuwu@unsrat.ac.id.

Lawrence J. L. Lumingas, Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115, North Sulawesi, Indonesia, e-mail: lillumingas@vahoo.com

Nego E. Bataragoa, Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115, North Sulawesi, Indonesia, e-mail: nebgoa@unsrat.ac.id

Silvester B. Pratasik, Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115, North Sulawesi, Indonesia, e-mail: spjong07@yahoo.com Frans F. Tilaar, Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115,

North Sulawesi, Indonesia, e-mail: fftilaar@unsrat.co.id

Meiske S. Salaki, Faculty of Fisheries and Marine Science, Sam Ratulangi University, Jl. Kampus Bahu, Manado-95115, North Sulawesi, Indonesia, e-mail: mssalaki@unsrat.ac.id

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

How to cite this article:

Rondonuwu A. B., Lumingas L. J. L., Bataragoa N. E., Pratasik S. B., Tilaar F. F., Salaki M. S., 2020 Mitochondrial CO1 sequences of banggai cardinal fish (BCF) from Lembeh Strait, North Sulawesi, Indonesia. AACL Bioflux 13(2):xxx-xxx.